

Article



⊘ Cite This: ACS Omega 2018, 3, 15182–15192

http://pubs.acs.org/journal/acsodf

Versatile Synthetic Approach for Selective Diversification of Bicyclic **Aza-Diketopiperazines**

Florent Péron, Stéphanie Riché, Brigitte Lesur, Marcel Hibert, Philippe Breton, Jean-Marie Fourquez, * Nicolas Girard, *, † 6 and Dominique Bonnet *, † 6

Supporting Information

ABSTRACT: Herein, we report a convenient synthesis of unprecedented aza-diketopiperazines (aza-DKPs). The strategy is based on selective diversification of bicyclic aza-DKP scaffolds by click reaction, N-acylation, and/or N-alkylation. These scaffolds containing either azido or amino groups were obtained by a key Rh(I)-catalyzed hydroformylative cyclohydrocarbonylation reaction of allyl-substituted aza-DKP. The methodology is readily amenable to the parallel synthesis of original aza-DKPs to enlarge the chemical diversity of aza-heterocycles.

■ INTRODUCTION

Small heterocyclic scaffolds are of central importance in the development of novel pharmacological probes and/or drug candidates. However, a recent literature analysis over the last 30 years has shown that an average of only five new ring systems per year have emerged.2 Therefore, to enlarge the chemical diversity of scaffolds, we decided to design and to synthesize novel aza-diketopiperazines (aza-DKPs) with potential applications in medicinal chemistry. By contrast to the well-known 2,5-diketopiperazines (2,5-DKPs),³ aza-DKPs have been much less exploited and few synthetic approaches have been reported to date. 4-10 Recently, we have shown that aza-DKPs were obtained following a deprotection/cyclization process with a higher yield than 2,5-DKPs. 10 In addition, the replacement of one C α -stereogenic center by a nitrogen atom resulted in a significant improvement of both the water solubility and the microsomal stability of this scaffold as compared to the 2,5-DKP analogue. 10 We have also described more complex aza-DKPs readily obtained following a novel domino cyclohydrocarbonylation (CHC)/addition process.9 This methodology enabled the rapid and efficient synthesis of novel three-dimensional (3D)-shaped bicyclic and tricyclic aza-DKP scaffolds containing sp³-hybridized carbon atoms and controlled stereocenters. Moreover, the analysis of their 3D molecular descriptors and "drug-likeness" properties highlighted not only their originality in the chemical space of azaheterocycles but also their higher likelihood of successfully progressing through the hit-to-lead process.9b

Herein, we report the synthesis of novel 6,6-bicyclic aza-DKPs showing two points of diversification at N-2 and C-3 positions of the scaffold (Figure 1). Cyclic systems fused to a 2,5-DKP showed interesting biological activities. 11-13 The ability to place the different substituents in a structurally rigid three-dimensional arrangement should allow for increased selectivity toward biological targets. The N-2 and C-3 positions were selected as diversification points, as they are strategic sites of functionalization for 2,5-DKPs. For instance, the N-2 position has been used as an anchor for the design of conjugated molecules between integrin ligands and paclitaxel for the synthesis of antitumor agents. ¹⁴ In some 2,5-DKPs-based compounds such as chaetocin A¹⁵ or E- and N-cadherin inhibitors, 16 the C-3 position is functionalized by an amido or alcohol groups. Therefore, in the strategy described herein, we have studied the possibility to introduce in this position both hydroxy- or amino-methyl groups as a starting point for the elongation of an O-alkyl or peptidic chains. The aminomethyl moiety was readily obtained from an azide derivative, which also allowed the formation of triazole rings. The latter are appealing in medicinal chemistry because they have been described as mimics of transoid amide bond which, like the aza-DKP backbone, allows to increase the metabolic stability of the synthetized molecules while maintaining biological features.17

Received: July 24, 2018 Accepted: October 25, 2018 Published: November 9, 2018



15182

[†]Laboratoire d'Innovation Thérapeutique, Labex MEDALIS, Faculté de Pharmacie, UMR7200 CNRS/Université de Strasbourg, 74 route du Rhin, 67412 Illkirch, France

[‡]Institut de Recherches Servier, 125 Chemin de Ronde, 78290 Croissy-Sur-Seine, France

Figure 1. General structure of novel bicyclic aza-DKPs and selected examples of known biologically active 2,5-DKPs.

Thereby, we herein report an unprecedented strategy for selective diversification of aza-DKPs to rapidly access mono- or disubstituted aza-DKPs. The strategy relies on the preparation of two aza-DKP bicyclic scaffolds encompassing at C-3 position either a hydroxymethyl (5) or an azidomethyl (8) moiety (Figure 2). Thereafter, subsequent functionalization of

Figure 2. Design of aza-DKP scaffolds 5 and 8 ready to functionalize.

these two scaffolds was evaluated and allowed access to N-acyl-, triazole-, or amido-substituted aza-DKPs. To introduce a

second point of diversification, the alkylation at N-2 was also investigated, leading to novel difunctionalized aza-DKPs. We anticipate that the strategy described herein will be useful to facilitate the parallel synthesis of complex aza-DKPs to enrich the chemical diversity of the existing chemical libraries.

■ RESULTS AND DISCUSSION

To access scaffolds 5 and 8 displaying two sites of potential derivatization, we envisioned a straightforward synthetic approach starting from N-protected-O-benzyl-L-serine methyl ester (Figure 2). The key step of the process is based on the preparation of the allyl-substituted aza-DKP precursor⁸ enabling the straightforward synthesis of the desired bicyclic aza-DKP scaffolds by cyclohydrocarbonylation reaction.⁹

Bicyclic Alcohol Functionalized Aza-DKP 5. To readily access this scaffold, the α -amino group of the commercially available O-benzyl-L-serine methyl ester was protected with the acid-sensitive p-methoxybenzyl (PMB) protecting group

Scheme 1. Preparation of Aza-DKP Scaffolds 5 and 8

through a reductive amination in the presence of pmethoxybenzaldehyde leading to alkylated amino ester 1 in 91% yield (Scheme 1). The latter was then reacted with bis(trichloromethyl)-carbonate and allyl t-butyl carbazate 2, obtained in one step from commercially available t-butyl carbazate, to provide semicarbazide intermediate. The subsequent N-Boc deprotection and cyclization were then attempted by treating crude semicarbazide with trifluoroacetic acid (TFA)/H₂O (95:5). Unfortunately, in these conditions, only the products arising from the simultaneous N-Boc and N-PMB deprotections were observed without ulterior cyclization. This result is consistent with our previous findings that demonstrated the necessity of starting from N-alkylated linear semicarbazide to initiate the cyclization toward aza-DKP.8 Similar result was obtained in diluted acidic conditions (5% of TFA in dichloromethane (DCM)), attesting the highly acidsensitive character of the PMB-protecting group. To circumvent this issue, a boiling-water-catalyzed neutral N-Boc cleavage method was applied. 18 The crude semicarbazide was dissolved in a mixture H₂O/1,4-dioxane (6:1) and heated to reflux for 18 h. Under these conditions, N-Boc group was removed selectively and the cyclization occurred spontaneously to provide aza-DKP 3 in 67% overall yield (two steps from 1).

The allyl derivative 3 was then submitted to a Rh(I)catalyzed cyclohydrocarbonylation (CHC) process to readily access bicyclic aza-DKP 4. As previously reported, the CHC was performed under classical conditions, i.e., the use of syngas (H₂/CO) in the presence of Rh(I) catalyst, BiPhePhos, and CSA in tetrahydrofuran (THF). However, in these conditions, a mixture of the desired bicyclic derivative 4 and the hydroxylated intermediate at C-6 position were obtained. Then, to facilitate the purification and to increase the yield of the reaction, the subsequent conversion of the hydroxyl derivative into aza-DKP 4 was attempted. However, regardless of the time, the temperature or the nature of the acid (CSA or PPTS), the full conversion of alcohol intermediate into aza-DKP 4 was never observed. To circumvent this limitation, we then envisaged a two-step process that involved the methoxylated derivative at the C-6 position and its subsequent elimination under acidic condition to provide the desired enamide 4. Therefore, the Rh(I)-catalyzed cyclohydrocarbonylation of aza-DKP 3 was achieved in MeOH/THF (10:1), leading to the methoxylated derivative at the C-6 position, which was directly subjected to acidic conditions (PPTS) to give exclusively the enamide 4 in 71% yield. Finally, the simultaneous reduction of the alkene and the removal of the benzyl-protecting group were performed by catalytic hydrogenolysis of 4 in the presence of Pearlman's catalyst. Thereby, scaffold 5 was obtained in excellent yield (95%), ready to be further functionalized.

Bicyclic Azido Functionalized Aza-DKP 8. To design original aza-DKPs via both click chemistry and reduction/N-acylation sequence from an azido derivative, aza-DKP scaffold 8 was efficiently obtained in a three-step process starting from bicyclic compound 5 (Scheme 1). The primary alcohol was first converted into the tosylate 6 followed by nucleophilic substitution in the presence of NaN₃ providing azido 7 in excellent yield (93%). The PMB group was then easily removed from 7 by heating at 40 °C in TFA/H₂O/TIS (95:2.5:2.5) to give the deprotected aza-DKP 8 in good yield (79%), ready to react via a copper-catalyzed azide—alkyne cycloaddition.

The efficient and robust process herein developed led to the synthesis of both scaffolds 5 and 8 in 41% overall yield (five steps) and 28% overall yield (eight steps), respectively. Noteworthy, this process was readily amenable to gram-scale production of both platforms for their subsequent functionalization.

Attempts To Functionalize Aza-DKP Scaffold 5. We then turned our attention to the diversification of both scaffolds 5 and 8. First, we examined the reactivity of primary alcohol 5 toward classical O-alkylation conditions (Scheme 2).

Scheme 2. Synthesis of Dehydroalanine-Containing Aza-DKP 9 from Aza-DKP 5 and 6

The use of NaH or inorganic bases (K_2CO_3 , Cs_2CO_3) in the presence of alkylating agent (allyl bromide or iodomethyl-cyclopentane) failed to provide the desired alkylated analogues. Regardless of the tested conditions, only the dehydration reaction was observed, leading to alkene 9 quasi-quantitatively. To avoid dehydration reaction, the alkylation of 5 was tested under base-free conditions by employing silver salts (Ag_2O) and alkyl halide in DCM. However, alcohol 5 proved to be unreactive toward these conditions (results not shown).

The alkylation was then envisaged following a Mitsunobu reaction. Compound 5 was reacted with PPh₃ and DIAD in the presence of 4-fluorophenol in THF. Unfortunately, as observed above, alkene 9 was quantitatively obtained. Finally, we tried an alternative strategy through tosylate derivative 6, obtained during the synthesis of scaffold 8. However, the reaction of 4-methoxyl phenol in the presence of NaH or Cs_2CO_3 in dimethylformamide (DMF) led exclusively to the dehydroalanine derivative 9. Taken together, these results demonstrate that the O-alkylation of serine system in the presence of a strong base or via alcohol activation remains difficult due to the high propensity of the α -proton (H-3) to undergo elimination. However, it is noteworthy that dehydroalanine-containing aza-DKP 9 could be a useful substrate for further functionalization including Michael addition as well as chemoselective ligation. 21

Derivatization of Aza-DKP Scaffold 8 via Click Chemistry. Owing to the difficulty to functionalize platform 5, we then focused our attention on the derivatization of aza-DKP scaffold 8 under copper(I)-catalyzed azide—alkyne cycloaddition conditions (CuAAC) to introduce a triazole moiety as a bioisostere of the amide bond (Scheme 3).¹⁷ Treatment of azido 8 with CuSO₄, sodium ascorbate, and phenylacetylene in THF/H₂O (1:0.8) at 25 °C for 4 h led to triazole-substituted aza-DKP **10a** in excellent yield (95%). This reaction was also successfully applied to other terminal alkynes substituted by heteroaryl group including thiophene and pyridine providing triazoles **10b** and **10c** in 95 and 76% yields, respectively.

Derivatization of Aza-DKP Scaffold 8 via Reduction and Subsequent N-Acylation. Taking advantage of the azido moiety, a first functionalization was envisaged following a two-step reduction/acylation process (Scheme 3). Therefore, azide 8 was submitted to catalytic hydrogenation in the

Scheme 3. Functionalization of Aza-DKP Scaffold 8

^a Isolated yields. ^b Diastereomeric ratios were determined by HPLC analysis of crude reaction mixtures.

presence of H₂/Pd in MeOH to provide amine intermediate quantitatively, which was straight way engaged in the Nacylation step after a simple filtration to remove the catalyst. Thereby, the primary amine was easily coupled with acyl chloride bearing aromatic and aliphatic groups affording amides 11a-c with yields ranging from 77 to 93%. To enrich the chemical diversity around bicyclic aza-DKPs, commercially available amino acid building blocks including Boc-(L)- and Boc-(D)-phenylalanine were also introduced via acylation under classical peptidic coupling conditions to provide diastereomers 11d and 11e in good yields (84 and 96%, respectively). These examples supplied information on the potential epimerization at the C-3 position all along the synthesis of azido 8, as only diastereomers 11d and 11e were obtained with only slight erosion of enantioselectivity (dr ratio 94:6 and 93:7, respectively). Finally, to envision the functionalization at N-2 position before the acylation of the primary amino group, the latter was efficiently protected by tert-butoxycarbonyl group in the presence of di-tert-butyl dicarbonate (11f, 94%).

N-Alkylation of Monofunctionalized Aza-DKP 11a. Aiming to access difunctionalized scaffolds, the reactivity of free nitrogen-containing aza-DKP (N-2) toward N-alkylation conditions was then explored starting from N-acylated compound 11a. Various bases including K2CO3, Cs2CO3, t-BuOK, or BEMP were evaluated in the presence of propargyl bromide in DMF and using tetrabutylammonium iodide (TBAI). As previously reported by Luthman et al. for the Nalkylation of free nitrogen-containing 2,5-DKPs, best conditions were obtained using the sterically hindered iminophosporane base (i.e., BEMP).²² Thereby, after 4 h at room temperature, the propargylic derivative 12a was isolated in 75% vield (Scheme 4). To evaluate the scope and limitation of the reaction, these conditions were then applied to diverse alkylating agents (R group, Scheme 4). Compared to the more reactive propargyl bromide, the alkylation of aza-DKP 11a with *n*-butyl bromide (12b) and Boc-aminopropyl bromide (12c) was sluggish, with only a partial conversion

Scheme 4. Access to Disubstituted Aza-DKPs by N-Alkylation of Scaffold 11a

 a Isolated yields. b Conversion of **10a** determined by HPLC is given in parentheses. c 3 equiv. of BEMP and 3 equiv. of alkylating agents were added after 6 hours.

of 11a in a nearly 1:1 ratio after 6 h at room temperature for both alkylating agents. Whereas the extension of reaction times or heating did not improve the conversion of 11a, the addition of 3 equiv of base and alkyl bromide proved to be beneficial and led to compounds 12b and 12c in 62 and 60% yields, respectively. Nevertheless, the reaction performed in the presence of sterically demanding isobutyl bromide led to a lower yield after 48 h regardless of the amount of base and alkyl bromide (12d, 10%). This low-yield alkylation can be ascribed to the steric hindrance of isobutyl bromide along with its propensity to undergo dehydrohalogenation (—HX) in the presence of base. Finally, the 4-fluorobenzyl group was introduced efficiently under the same conditions without the need of additional base or alkylating agent providing compound 12e in excellent yield (89%).

Overall, this methodology allows access to novel disubstituted bicyclic aza-DKPs bearing functionalities such as propargyl or amino groups able to react with various commercial building blocks to further extend the chemical diversity of this scaffold. Noteworthy, this is a rare example of alkylation of secondary amine containing aza-urea moiety.²³

CONCLUSIONS

In this article, we described an efficient and versatile approach for selective diversification of mono- and disubstituted bicyclic aza-DKPs to enlarge the chemical diversity accessible around this scaffold. The synthetic route involved azido scaffold 8 obtained in eight steps in 28% overall yield from the commercially available O-benzyl-L-serine methyl ester. Then, we took advantage of the double reactivity of the azido moiety to access various unprecedented aza-heterocycles such as (1) triazole-containing aza-DKPs and (2) acylated aza-DKPs obtained after the reduction of the azido function and subsequent N-acylation. To prepare disubstituted scaffolds, we have evaluated the possibility to alkylate the benzoylated intermediate. The use of sterically hindered iminophosporane base allowed us to generate various disubstituted bicyclic aza-DKPs. All attempts to alkylate scaffold 5 having led to the formation of the dehydroalanine 9, it will be interesting to evaluate the reactivity of this new compound with various nucleophiles.

EXPERIMENTAL SECTION

General Methods. Reagents were obtained from commercial sources and used without any further purification. Thinlayer chromatography was performed on silica gel 60 F₂₅₄ plates. (Acetylacetonato)dicarbonylrhodium(I) and dry solvents were purchased from Sigma-Aldrich. BiPhePhos was prepared as reported previously.²⁴ All the experiments were performed under argon atmosphere except where otherwise noted. Hydroformylation was performed in a reactor from Equilabo using 1:1 H₂/CO supplied by Airgas, Inc. Flash chromatography was performed on silica gel (60 Å, particle size $40-63 \mu m$). Purification by semipreparative HPLC was performed on RP-18 (25-40 µm, Merck) prepacked columns on a PLC 2020 apparatus from Gilson. ¹H, ¹³C, and ¹⁹F NMR spectra were recorded, respectively, at 400, 100, and 376 MHz with a Bruker Advance 400 spectrometer. Conditions are specified for each spectrum (temperature 25 °C unless specified). Chemical shifts are reported in parts per million (ppm) relative to residual solvent and coupling constants (J) are reported in hertz (Hz). Signals are described as s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet), brs (broad singlet), brd (broad doublet), brq (broad quadruplet), or a combination of the above. Specific rotations were measured on a Perkin-Elmer polarimeter using a 10 cm cell with a sodium lamp at 589 nm. High-resolution mass spectra were obtained on an Agilent Technologie 6520 Accurate-Mass Q-TOF LC/ MC apparatus using electrospray ionization mode and time-offlight analyzer (ESI-TOF).

General Procedure A. To a degassed solution of 8 (1 equiv) in a mixture THF/H₂O (1:0.8, 15 mL mmol⁻¹) were successively added heteroarylacetylene (1.1 equiv), sodium ascorbate (0.8 equiv), and CuSO₄·5H₂O (0.3 equiv) at room temperature under argon. The resulting solution was degassed for 5 min and then stirred at room temperature for 4 h under argon. This mixture was then concentrated in vacuo and the

resulting residue was partitioned between EtOAc (125 mL mmol⁻¹) and saturated aqueous NH₄Cl solution (62.5 mL mmol⁻¹). The aqueous layer was extracted three times with EtOAc (125 mL mmol⁻¹), the combined organic layers were washed with brine (250 mL mmol⁻¹), dried over anhydrous Na₂SO₄, and concentrated in vacuo. The crude mixture was purified by silica gel chromatography (gradient 50–95% of EtOAc in n-pentane).

General Procedure B. To a solution of 8 (1 equiv) in dry MeOH (11.2 mL mmol⁻¹) was added 10 wt % Pd on carbon under argon. The resulting mixture was stirred at room temperature under a hydrogen atmosphere for 3 h. The reaction mixture was filtered over a pad of Celite and washed twice with MeOH (22.4 mL mmol⁻¹). The filtrate was concentrated in vacuo and the resulting residue was used as such in the next step. To a solution of the crude mixture in dry DCM (9.0 mL mmol⁻¹) were added successively Et₃N (1.2 equiv) and acid chloride (1.1 equiv) dropwise at 0 °C under argon. The reaction mixture was stirred at 0 °C for 1 h under argon. The resulting mixture was diluted with DCM (44.8 mL mmol⁻¹) and H₂O (22.4 mL mmol⁻¹) and the aqueous layer was extracted three times with DCM (44.8 mL mmol⁻¹). The combined organic layers were washed with brine (112 mL mmol⁻¹), dried over anhydrous Na₂SO₄, and concentrated in vacuo. The crude mixture was purified by silica gel chromatography (gradient 0-3% of MeOH in DCM).

General Procedure C. To a solution of 8 (1 equiv) in dry MeOH (11.2 mL mmol⁻¹) was added 10 wt % Pd on carbon under argon. The resulting mixture was stirred at room temperature under a hydrogen atmosphere for 3 h. The reaction mixture was filtered over a pad of Celite, which was washed twice with MeOH (22.4 mL mmol⁻¹). The filtrate was concentrated in vacuo and the resulting residue was used as such in the next step. To a solution of the crude mixture in dry DMF (11 mL mmol⁻¹) were added successively PyBOP (1.1 equiv), Boc-amino acid (1.1 equiv), and N,N-diisopropylethylamine (DIEA, 4 equiv) at room temperature under argon. The reaction mixture was stirred for 2 h at this temperature under argon and then concentrated in vacuo. The resulting residue was diluted with DCM (22.4 mL mmol⁻¹) and H₂O (11.2 mL mmol⁻¹) and the aqueous layer was extracted three times with DCM (22.4 mL mmol⁻¹). The combined organic layers were washed with brine (56 mL mmol⁻¹), dried over anhydrous Na₂SO₄, and concentrated in vacuo. The crude mixture was purified by silica gel chromatography (gradient 0-3% of MeOH in DCM).

General Procedure D. To a solution of 11a (1 equiv) in dry DMF (10.1 mL mmol⁻¹) were added successively BEMP (3 equiv), TBAI (3 equiv), and an alkylating agent (5 equiv) at room temperature under argon. The resulting mixture was stirred for 3–48 h at this temperature under argon. The resulting solution was then diluted with $\rm H_2O$ (50 mL mmol⁻¹) and extracted three times with EtOAc (150 mL mmol⁻¹). The combined organic layers were washed with brine (300 mL mmol⁻¹), dried over $\rm Na_2SO_4$, and concentrated in vacuo. The crude mixture was purified by silica gel chromatography (gradient 0–80% of EtOAc in n-pentane).

O-Benzyl-N-(4-methoxybenzyl)-L-serine Methyl Ester (1).²⁵ To a solution of O-benzyl-L-serine methyl ester hydrochloride (2.70 g, 10.9 mmol, 1 equiv) in anhydrous methanol (95 mL) was added triethylamine (1.53 mL, 10.9 mmol, 1 equiv) at room temperature under argon. The reaction mixture was stirred for 10 min and p-anisaldehyde (1.20 mL, 9.9 mmol, 0.9

equiv) was then added. After stirring for 1 h, the solution was cooled to 0 °C and then acetic acid (1.26 mL, 16.5 mmol, 2 equiv) and NaBH₃CN (1.04 g, 16.5 mmol, 1.5 equiv) were successively added. The resulting solution was stirred at 0-5 °C for 1 h and then at room temperature for a further 18 h under argon. Solid NaHCO₃ (2.77 g, 32.9 mmol, 3 equiv) was added and the solvent was removed under reduced pressure. The resulting white residue was partitioned between DCM (170 mL) and H₂O (80 mL) and the aqueous layer was extracted with DCM (2 × 170 mL). The combined organic layers were washed with brine (200 mL), dried over anhydrous Na₂SO₄, and concentrated in vacuo. The crude residue was purified by silica gel chromatography (0-20% of EtOAc in npentane) affording 1 as a colorless oil (2.95 g, 8.96 mmol, 91%). $R_f = 0.28$ (*n*-pentane/EtOAc, 3:2); $[\alpha]_D^{20} = -8.53$ (*c* 0.23, MeOH); ¹H NMR (400 MHz, CDCl₃) δ 7.36–7.26 (m, 5H), 7.25 (d, J = 8.6 Hz, 2H), 6.86 (d, J = 8.6 Hz, 2H), 4.54 (d, J = 12.2 Hz, 1H), 4.49 (d, J = 12.2 Hz, 1H), 3.83 (d, J = 12.2 Hz, 1H)12.8 Hz, 1H), 3.80 (s, 3H), 3.74 (s, 3H), 3.72-3.65 (m, 2H), 3.65 (d, J = 12.8 Hz, 1H), 3.51 (t, J = 4.9 Hz, 1H), 2.05 (brs,1H); 13 C NMR (100 MHz, CDCl₃) δ 173.6, 158.7, 137.9, 131.7, 129.5 (2C), 128.3 (2C), 127.6, 127.6 (2C), 113.8 (2C), 73.2, 71.0, 60.4, 55.2, 51.9, 51.4; HRMS (ESI-TOF): calcd for $C_{19}H_{24}NO_4 [M + H]^+ 330.1705$, found 330.1704.

tert-Butyl 2-Allylhydrazinecarboxylate (2).9a To a dry 100 mL flask were added tert-butyl carbazate (4.22 g, 31.9 mmol, 3 equiv), potassium carbonate (1.47 g, 10.6 mmol, 1 equiv), and anhydrous THF/DMF (9:1, 30.7 mL). The suspension was heated to 80 °C. A solution of allyl bromide (920 µL, 10.6 mmol, 1 equiv) in a mixture THF/DMF (9:1, 4 mL) was added over 3 h and the mixture was stirred overnight at 80 °C. The mixture was then concentrated in vacuo to a slurry and the crude was diluted with H₂O/EtOAc (1:1, 400 mL). The aqueous layer was extracted with EtOAc (2×200 mL). The combined organic layers were washed with brine (300 mL), dried over anhydrous Na₂SO₄, and concentrated in vacuo. The crude product was purified by silica gel chromatography (0-25% of EtOAc in n-heptane) yielding 2 (1.21 g, 7.02 mmol, 66%) as a colorless oil. $R_f = 0.26$ (*n*-heptane/EtOAc, 7:3); ¹H NMR (400 MHz, CDCl₃) δ 6.17 (brs, 1H), 5.84 (ddt, J = 16.6, 10.4, 6.3 Hz, 1H), 5.22 (brd, *J* = 17.1 Hz, 1H), 5.16 (brd, *J* = 10.2 Hz, 1H), 4.00 (brs, 1H), 3.46 (d, J = 6.3 Hz, 2H), 1.46 (s, 9H); 13 C NMR (100 MHz, CDCl₃) δ 156.7, 134.2, 118.0, 80.4, 54.6, 28.3 (3C).

(S)-2-Allyl-5-((benzyloxy)methyl)-4-(4-methoxybenzyl)-1,2,4-triazinane-3,6-dione (3). To a solution of triphosgene (0.315 g, 1.06 mmol, 0.35 equiv) in anhydrous THF (20 mL), under argon, was added dropwise (over 10 min) a solution of 1 (1.00 g, 3.04 mmol, 1 equiv) and DIEA (0.578 mL, 3.34 mmol, 1.1 equiv) in a mixture of anhydrous THF/DCM (2:1, 12 mL). The resulting mixture was stirred at room temperature for 25 min. A solution of 2 (0.549 g, 3.19 mmol, 1.05 equiv) and DIEA (0.578 mL, 3.34 mmol, 1.1 equiv) in anhydrous THF (6 mL) was then added dropwise and the mixture was heated at 40 °C overnight. The mixture was evaporated in vacuo, partitioned between EtOAc (150 mL) and H₂O (50 mL), and the aqueous layer was re-extracted twice with EtOAc $(2 \times 150 \text{ mL})$. The combined organic layers were washed successively with an aqueous solution of citric acid (5%, 100 mL), a saturated solution of NaHCO3 (100 mL) and with brine (100 mL), dried over anhydrous Na₂SO₄, and concentrated in vacuo. In a sealed flask, the crude residue was dissolved in 1,4-dioxane (24 mL) and H₂O (140 mL) was

added. The mixture was heated overnight at 120 °C and evaporated under reduced pressure. The crude residue was diluted with EtOAc/H₂O (2:1, 150 mL) and the aqueous layer was extracted twice with EtOAc (2 \times 75 mL). The combined organic layers were washed with brine (150 mL), dried over anhydrous Na₂SO₄, and concentrated in vacuo. The crude mixture was purified by silica gel chromatography (gradient 0-50% of EtOAc in *n*-pentane) leading to 3 (0.800 g, 2.02 mmol, 67%) as a colorless oil. $R_f = 0.27$ (n-pentane/EtOAc, 1:1); $[\alpha]_D^{20} = +28.6$ (c 0.20, MeOH); ¹H NMR (400 MHz, CDCl₃) δ 8.74 (brs, 1H), 7.38–7.29 (m, 5H), 7.19 (d, J = 8.5 Hz, 2H), 6.85 (d, J = 8.5 Hz, 2H), 5.75 (dddd, J = 17.2, 10.2, 7.2, 5.7 Hz, 1H), 5.26-5.18 (m, 2H), 4.94 (d, J = 14.9 Hz, 1H), 4.49 (d, J = 11.8 Hz, 1H), 4.43 (d, J = 11.8 Hz, 1H), 4.25 (dd, J = 11.8 Hz, 1H)15.3, 5.4 Hz, 1H), 4.07 (d, J = 14.9 Hz, 1H), 3.85 (t, J = 3.5Hz, 1H), 3.83-3.78 (m, 4H), 3.66 (dd, J = 9.9, 3.4 Hz, 1H), 3.55 (dd, J = 9.9, 3.4 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 164.1, 159.4, 154.2, 137.1, 131.6, 129.7 (2C), 128.5 (2C), 128.1, 128.0, 127.9 (2C), 120.2, 114.2 (2C), 73.6, 67.1, 58.9, 55.3, 52.1, 48.6; HRMS (ESI-TOF): calcd for C₂₂H₂₆N₃O₄ [M + H]+ 396.1923, found 396.1912.

(S)-3-((Benzyloxy)methyl)-2-(4-methoxybenzyl)-2,3,8,9tetrahydropyridazino[1,2-a][1,2,4]-triazine-1,4-dione (4). In a Schlenk glassware under argon were added Rh(CO)₂(acac) (13 mg, 0.051 mmol, 0.02 equiv) and anhydrous THF (4 mL). BiPhePhos (0.119 g, 0.152 mmol, 0.06 equiv) was then added at room temperature and this mixture was stirred at this temperature for 10 min. The resulting solution was introduced under argon into a stainless steel autoclave containing a solution of 3 (1.00 g, 2.53 mmol, 1 equiv) and camphor sulfonic acid (0.176 g, 0.759 mmol, 0.3 equiv) in anhydrous methanol (40 mL). The reactor was purged three times with H_2/CO (1:1, 5 bars) and filled with H_2/CO (1:1, 5 bar). The reactor was heated to 70 °C and stirred for 16 h. The reactor was then cooled to room temperature and vented to atmospheric pressure. The reaction mixture was quenched with solid NaHCO₃ (0.425 g, 5.06 mmol, 2 equiv) and then evaporated in vacuo. The resulting residue was partitioned between EtOAc (50 mL) and H₂O (25 mL) and the aqueous layer was re-extracted twice with EtOAc (2 \times 40 mL). The combined organic layers were washed with brine (50 mL), dried over anhydrous Na₂SO₄, and concentrated in vacuo. To a solution of the crude residue in anhydrous toluene (45 mL) was added pyridinium p-toluenesulfonate (0.318 g, 1.26 mmol, 0.5 equiv) at room temperature under argon. The reaction mixture was heated at 110 °C for 6 h. The resulting mixture was then cooled to room temperature, quenched with solid NaHCO₃ (0.425 g, 5.06 mmol, 2 equiv), and evaporated in vacuo. The resulting residue was partitioned between EtOAc (50 mL) and H₂O (25 mL) and the aqueous layer was extracted with EtOAc (2×50 mL). The combined organic layers were washed with brine (50 mL), dried over anhydrous Na₂SO₄, and concentrated in vacuo. The crude mixture was purified by silica gel chromatography (gradient 0-25% of EtOAc in n-pentane) leading to 4 as a colorless oil (0.732 g, 1.79 mmol, 71%). $R_f = 0.47$ (*n*-pentane/EtOAc, 1:1); $[\alpha]_D^{20} =$ +118.8 (c 0.18, MeOH); ¹H NMR (400 MHz, CDCl₃) δ 7.37-7.27 (m, 5H), 7.20 (d, J = 8.6 Hz, 2H), 7.06 (dd, J = 8.4, 1.5 Hz, 1H), 6.84 (d, J = 8.6 Hz, 2H), 5.29 (brt, J = 6.4 Hz, 1H), 4.89 (d, J = 14.8 Hz, 1H), 4.51-4.46 (m, 1H), 4.46 (d, J= 11.6 Hz, 1H), 4.41 (d, J = 11.6 Hz, 1H), 4.12 (d, J = 14.8)Hz, 1H), 3.88 (t, J = 3.2 Hz, 1H), 3.79 (s, 3H), 3.67 (dd, J =9.9, 3.3 Hz, 1H), 3.56 (dd, *J* = 9.9, 3.3 Hz, 1H), 2.83 (ddd, *J* =

12.7, 11.4, 3.8 Hz, 1H), 2.51–2.41 (m, 1H), 2.09 (dt, J = 17.4, 4.8 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 160.2, 159.4, 153.6, 137.1, 129.7 (2C), 128.4 (2C), 128.1, 127.9, 127.8 (2C), 119.8, 114.2 (2C), 108.3, 73.6, 67.6, 58.5, 55.2, 48.4, 41.5, 22.9; HRMS (ESI-TOF): calcd for $C_{23}H_{26}N_3O_4$ [M + H]⁺ 408.1923, found 408.1910.

(S)-3-(Hydroxymethyl)-2-(4-methoxybenzyl)hexahydropyridazino[1,2-a][1,2,4]triazine-1,4-dione (5). To a solution of 4 (0.700 g, 1.72 mmol, 1 equiv) in EtOAc (50 mL) was added 20 wt % Pd(OH)2 on carbon (140 mg) under argon. The resulting solution was stirred at room temperature under an atmosphere of hydrogen for 16 h. The reaction mixture was filtered over a pad of Celite, which was washed with EtOAc (2 \times 50 mL). The filtrate was concentrated in vacuo and the resulting residue was purified by silica gel chromatography (gradient 50-100% of EtOAc in *n*-pentane) leading to 5 as a white amorphous solid (0.540 g, 1.69 mmol, 95%). $R_f = 0.16$ (EtOAc); $[\alpha]_D^{20} = +34.3$ (c 0.15, MeOH); ¹H NMR (400 MHz, CDCl₃) δ 7.20 (d, J = 8.6 Hz, 2H), 6.86 (d, J= 8.6 Hz, 2H), 4.91 (d, J = 14.8 Hz, 1H), 4.55 (brdd, J = 12.7,3.5 Hz, 1H), 4.27 (brd, J = 12.0 Hz, 1H), 4.08 (d, J = 14.8 Hz, 1H), 3.79 (s, 3H), 3.78-3.71 (m, 3H), 2.99-2.85 (m, 3H), 1.82-1.74 (m, 3H), 1.61-1.48 (m, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 162.6, 159.4, 156.2, 129.7 (2C), 127.8, 114.3 (2C), 59.7, 59.6, 55.3, 48.2, 48.1, 42.6, 23.6, 23.3; HRMS (ESI-TOF): calcd for $C_{16}H_{22}N_3O_4$ [M + H]⁺ 320.1610, found

(S)-(2-(4-Methoxybenzyl)-1,4-dioxooctahydropyridazino-[1,2-a][1,2,4]triazin-3-yl)methyl-4-methyl-benzenesulfonate (6). To a solution of 5 (1.34 g, 4.19 mmol, 1 equiv) in anhydrous DCM (80 mL) were added triethylamine (2.34 mL, 16.8 mmol, 4 equiv) and p-toluenesulfonyl chloride (3.20 mg, 16.8 mmol, 4 equiv) at 0 °C under argon. The reaction mixture was warmed to room temperature and stirred for 24 h. The resulting mixture was diluted with DCM (100 mL) and H₂O (100 mL) and the aqueous layer was extracted with DCM (3 \times 120 mL). The combined organic layers were washed with brine (200 mL), dried over anhydrous Na₂SO₄, and concentrated in vacuo. The crude mixture was purified by silica gel chromatography (gradient 0-50% of EtOAc in *n*-pentane) leading to 6 as a white amorphous solid (1.87 g, 3.94 mmol, 94%). $R_f = 0.51$ (EtOAc); $[\alpha]_D^{20} = +35.3$ (c 0.36, MeOH); ¹H NMR (400 MHz, CDCl₃) δ 7.76 (d, J = 8.2 Hz, 2H), 7.39 (d, J= 8.2 Hz, 2H), 7.07 (d, J = 8.5 Hz, 2H), 6.79 (d, J = 8.5 Hz,2H), 4.76 (d, J = 14.8 Hz, 1H), 4.54 (dd, J = 12.7, 3.5 Hz, 1H), 4.36 (brd, J = 13.4 Hz, 1H), 4.11 (dd, J = 10.2, 2.3 Hz, 1H), 3.92 (dd, J = 10.2, 2.9 Hz, 1H), 3.85 (brt, J = 2.3 Hz, 1H), 3.78(s, 3H), 3.77 (d, J = 14.8 Hz, 1H), 2.94 (td, J = 12.3, 2.6 Hz, 1H), 2.85 (td, J = 12.7, 2.4 Hz, 1H), 2.48 (s, 3H), 1.84–1.73 (m, 3H), 1.60–1.49 (m, 1H); 13 C NMR (100 MHz, CDCl₃) δ 159.9, 159.6, 154.7, 145.5, 131.9, 130.0 (2C), 129.7 (2C), 128.3 (2C), 127.1, 114.5 (2C), 65.8, 57.0, 55.3, 48.1, 47.8, 42.9, 23.5, 23.2, 21.7; HRMS (ESI-TOF): calcd for $C_{23}H_{28}N_3O_6S$ [M + H]⁺ 474.1699, found 474.1698.

(S)-3-(Azidomethyl)-2-(4-methoxybenzyl)-hexahydropyridazino[1,2-a][1,2,4]triazine-1,4-dione (7). To a solution of 6 (1.075 g, 2.27 mmol, 1 equiv) in anhydrous DMF (30 mL) was added NaN₃ (0.267 g, 4.09 mmol, 1.8 equiv) at room temperature under argon. The reaction mixture was stirred at 40 °C for 6 h under argon, then quenched with saturated aqueous NaHCO₃ solution (5 mL), and concentrated in vacuo. The resulting residue was partitioned between EtOAc (75 mL) and H₂O (30 mL) and the aqueous layer was

extracted with EtOAc (3 \times 50 mL). The combined organic layers were washed with brine (120 mL), dried over anhydrous Na₂SO₄, and concentrated in vacuo. The crude mixture was purified by silica gel chromatography (gradient 0-50% of EtOAc in *n*-pentane), leading to 7 as a white amorphous solid (0.727 g, 2.11 mmol, 93%). $R_f = 0.24 \text{ (n-pentane/EtOAc}, 1:1);$ $[\alpha]_D^{20} = +25.9$ (c 0.21, MeOH); ¹H NMR (400 MHz, CDCl₃) δ 7.24 (d, J = 8.6 Hz, 2H), 6.87 (d, J = 8.6 Hz, 2H), 4.70 (d, J= 14.8 Hz, 1H), 4.55 (brdd, *J* = 12.7, 3.7 Hz, 1H), 4.42 (brd, *J* = 12.6 Hz, 1H), 4.26 (d, J = 14.8 Hz, 1H), 3.85 (t, J = 3.1 Hz, 1H), 3.80 (s, 3H), 3.56 (dd, J = 12.8, 2.6 Hz, 1H), 3.34 (dd, J = 12.8, 3.8 Hz, 1H), 2.99 (td, J = 12.3, 2.8 Hz, 1H), 2.86 (td, J= 12.8, 2.7 Hz, 1H), 1.85–1.72 (m, 3H), 1.62–1.51 (m, 1H); 13 C NMR (100 MHz, CDCl₃): δ 161.0, 159.6, 154.7, 129.8 (2C), 127.8, 114.4 (2C), 57.8, 55.3, 50.3, 48.8, 48.0, 42.8, 23.5, 23.2; HRMS (ESI-TOF): calcd for $C_{16}H_{21}N_6O_3$ [M + H] 345.1675, found 345.1662.

(S)-3-(Azidomethyl)hexahydropyridazino[1,2-a][1,2,4]triazine-1,4-dione (8). A solution of 7 (0.820 g, 2.38 mmol, 1 equiv) in a mixture TFA/H₂O/TIS (95:2.5:2.5, 8 mL) was stirred at 40 °C for 18 h and then concentrated in vacuo. The resulting residue was quenched with saturated aqueous NaHCO₃ solution until pH > 7 and diluted with EtOAc (50 mL). The aqueous layer was extracted with EtOAc (3 \times 50 mL). The combined organic layers were washed with brine (75 mL), dried over anhydrous Na2SO4, and concentrated in vacuo. The crude mixture was purified by silica gel chromatography (gradient 0-90% of EtOAc in *n*-pentane) leading to **8** as a white amorphous solid (0.419 g, 1.87 mmol, 79%). $R_f = 0.15$ (*n*-pentane/EtOAc, 1:4); $[\alpha]_D^{20} = -80.1$ (*c* 0.14, MeOH); ¹H NMR (400 MHz, CDCl₃) δ 6.44 (brs, 1H), 4.03-4.00 (m, 2H), 3.90-3.86 (m, 1H), 3.69 (d, J = 4.3 Hz, 2H), 3.48-3.43 (m, 2H), 1.79-1.65 (m, 4H); ¹³C NMR (100 MHz, CDCl₃): δ 161.5, 155.4, 53.8, 52.7, 46.3, 43.2, 23.3, 23.0; HRMS (ESI-TOF): calcd for $C_8H_{13}N_6O_2 [M + H]^+$ 225.1100, found 225.1094.

2-(4-Methoxybenzyl)-3-methylenehexahydropyridazino-[1,2-a][1,2,4]triazine-1,4-dione (9). To a solution of 5 (86 mg, 0.269 mmol, 1 equiv), 4-fluorophenol (31 mg, 0.269 mmol, 1 equiv), and triphenylphosphine (71 mg, 0.269 mmol, 1 equiv) in anhydrous THF (4 mL) was added diisopropyl azodicarboxylate (58 μ L, 0.296 mmol, 1.1 equiv) at 0 °C under argon. The resulting mixture was stirred at room temperature for 6 h under argon. The mixture was concentrated in vacuo, partitioned between EtOAc (30 mL), and a saturated solution of NaHCO₃ (30 mL). The organic layer was separated and washed successively with H₂O (20 mL) and brine (20 mL), dried over anhydrous Na₂SO₄, and concentrated in vacuo. The crude residue was purified by silica gel chromatography (gradient 0–40% of EtOAc in *n*-pentane), leading to 9 as a white amorphous solid (74 mg, 0.255 mmol, 91%). $R_f = 0.31$ (*n*-pentane/EtOAc, 1:1); ¹H NMR (400 MHz, $CDCl_3$) δ 7.18 (d, J = 8.7 Hz, 2H), 6.85 (d, J = 8.7 Hz, 2H), 5.48 (brd, J = 1.1 Hz, 1H), 4.85 (s, 2H), 4.68 (brd, J = 1.2 Hz, 1H), 3.88 (t, J = 5.7 Hz, 2H), 3.80 (t, J = 5.3 Hz, 2H), 3.78 (s, 3H), 1.88-1.83 (m, 2H), 1.79-1.75 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 158.9, 155.9, 151.2, 134.5, 128.1 (2C), 127.5, 114.1 (2C), 99.6, 55.2, 48.3, 47.7, 43.8, 23.4, 22.9; HRMS (ESI-TOF): calcd for $C_{16}H_{20}N_3O_3 [M + H]^+$ 302.1505, found 302.1503.

(S)-3-((4-Phenyl-1H-1,2,3-triazol-1-yl)methyl)-hexahydropyridazino[1,2-a][1,2,4]-triazine-1,4-dione (10a). Starting from 8 (18 mg, 0.080 mmol) and phenylacetylene (10

 μ L, 0.088 mmol), using general procedure A, **10a** was obtained as a white amorphous solid (25 mg, 0.077 mmol, 96%). R_f = 0.13 (DCM/MeOH, 9.8:0.2); $[\alpha]_D^{20}$ = -52.9 (c 0.11, MeOH); ¹H NMR (400 MHz, CDCl₃) δ 7.97 (s, 1H), 7.79 (d, J = 7.3 Hz, 2H), 7.40 (t, J = 7.4 Hz, 2H), 7.33 (t, J = 7.3 Hz, 1H), 6.84 (brs, 1H), 4.90 (dd, J = 14.4, 3.8 Hz, 1H), 4.76 (dd, J = 14.4, 5.6 Hz, 1H), 4.45 (brt, J = 4.3 Hz, 1H), 3.73–3.64 (m, 2H), 3.49–3.46 (m, 1H), 3.33–3.30 (m, 1H), 1.68–1.59 (m, 4H); ¹³C NMR (100 MHz, CDCl₃) δ 161.2, 154.6, 148.0, 130.2, 128.9 (2C), 128.3, 125.7 (2C), 121.4, 54.1, 50.8, 45.0, 43.5, 23.1, 22.8; HRMS (ESI-TOF): calcd for C₁₆H₁₉N₆O₂ [M + H]⁺ 327.1569, found 327.1561.

(S)-3-((4-(Thiophen-3-yl)-1H-1,2,3-triazol-1-yl)methyl)hexahydropyridazino[1,2-a]-[1,2,4]triazine-1,4-dione (10b). Starting from 8 (20 mg, 0.089 mmol) and 3-ethynylthiophene (10 μ L, 0.098 mmol), using general procedure A, 10b was obtained as a white amorphous solid (28 mg, 0.084 mmol, 95%). $R_f = 0.15$ (EtOAc); $[\alpha]_D^{20} = -48.5$ (c 0.10, MeOH); ¹H NMR (400 MHz, DMSO- d_6) δ 8.32 (s, 1H), 7.86 (d, J = 2.5Hz, 1H), 7.64 (dd, J = 4.8, 3.0 Hz, 1H), 7.56 (brd, J = 1.4 Hz, 1H), 7.51 (d, J = 5.0 Hz, 1H), 4.69 (dd, J = 14.3, 4.6 Hz, 1H), 4.60 (dd, J = 14.3, 4.8 Hz, 1H), 4.38-4.35 (m, 1H), 3.95 (dt, J = 14.3, 4.8 Hz, 1H), 4.38-4.35 (m, 1H), 3.95 (dt, J = 14.3, 4.8 Hz, 1H), 4.38-4.35 (m, 1H), 3.95 (dt, J = 14.3, 4.8 Hz, 1H), 4.38-4.35 (m, 1H), 3.95 (dt, J = 14.3, 4.8 Hz, 1H), 4.38-4.35 (m, 1H), 3.95 (dt, J = 14.3, 4.8 Hz, 1H), 4.38-4.35 (m, 1H), 3.95 (dt, J = 14.3, 4.8 Hz, 1H), 4.38-4.35 (m, 1H), 3.95 (dt, J = 14.3, 4.8 Hz, 1H), 4.38-4.35 (m, 1H), 4.= 12.6, 4.3 Hz, 1H), 3.61 (dt, J = 12.2, 4.2 Hz, 1H), 3.17 (ddd, J = 12.6, 10.1, 2.8 Hz, 1H), 2.76 (ddd, J = 12.4, 9.4, 3.1 Hz, 1H), 1.64-1.47 (m, 3H), 1.44-1.38 (m, 1H); ¹³C NMR (100 MHz, DMSO- d_6) δ 160.8, 153.4, 142.8, 131.8, 127.1, 125.8, 122.2, 120.9, 53.8, 50.9, 44.8, 42.5, 22.9, 22.5; HRMS (ESI-TOF): calcd for $C_{14}H_{17}N_6O_2S$ [M + H]⁺ 333.1134, found 333.1117.

(S)-3-((4-(Pyridin-3-yl)-1H-1,2,3-triazol-1-yl)methyl)hexahydropyridazino[1,2-a]-[1,2,4]triazine-1,4-dione (10c). Starting from 8 (20 mg, 0.089 mmol) and 3-ethynylpyridine (10 mg, 0.098 mmol), using general procedure A, 10c was obtained as a white amorphous solid (22 mg, 0.067 mmol, 76%). $R_f = 0.22$ (DCM/MeOH, 9.6:0.4); $[\alpha]_D^{20} = -46.8$ (c 0.10, MeOH); ¹H NMR (400 MHz, DMSO- d_6) δ 9.04 (d, I =1.8 Hz, 1H), 8.60 (s, 1H), 8.55 (d, J = 4.8 Hz, 1H), 8.20 (d, J = 4.8 Hz, 1H), 8. = 7.9 Hz, 1H, 7.57 (s, 1H), 7.49 (dd, I = 7.9, 4.8 Hz, 1H),4.73 (dd, J = 14.3, 4.8 Hz, 1H), 4.65 (dd, J = 14.3, 4.6 Hz, 1.00 Hz1H), 4.39 (brt, I = 4.5 Hz, 1H), 3.94 (dt, I = 12.6, 4.0 Hz, 1H), 3.62 (dt, I = 12.6, 4.5 Hz, 1H), 3.18 (ddd, I = 12.5, 10.1, 2.6Hz, 1H), 2.77 (ddd, I = 12.4, 9.4, 3.1 Hz, 1H), 1.64–1.40 (m, 4H); 13 C NMR (101 MHz, DMSO- d_6) δ 160.8, 153.4, 149.0, 146.4, 143.5, 132.5, 126.5, 124.0, 123.2, 53.8, 51.1, 44.7, 42.6, 22.9, 22.5; HRMS (ESI-TOF): calcd for $C_{15}H_{18}N_7O_2$ [M + H]⁺ 328.1522, found 328.1508.

(S)-N-((1,4-Dioxooctahydropyridazino[1,2-a][1,2,4]-triazin-3-yl)methyl)benzamide (11a). Starting from 8 (0.100 g, 0.446 mmol) and benzoyl chloride (57 μL, 0.488 mmol), using general procedure B, 11a was obtained as a white amorphous solid (0.125 g, 0.413 mmol, 93%). $R_f = 0.13$ (DCM/MeOH, 9.6:0.4); $[\alpha]_D^{20} = -32.7$ (c 0.09, MeOH); 1 H NMR (400 MHz, CDCl₃) δ 7.80 (d, J = 7.6 Hz, 2H), 7.50 (t, J = 7.5 Hz, 1H), 7.42 (t, J = 7.5 Hz, 2H), 7.19–7.17 (m, 1H), 6.41 (brs, 1H), 4.07–4.00 (m, 3H), 3.91 (brd, J = 12.4 Hz, 1H), 3.77–3.71 (m, 1H), 3.34–3.22 (m, 2H), 1.77–1.66 (m, 4H); 13 C NMR (100 MHz, CDCl₃) δ 168.5, 163.7, 155.7, 133.7, 131.8, 128.5 (2C), 127.2 (2C), 54.0, 44.5, 43.4, 39.8, 23.4, 22.9; HRMS (ESI-TOF): calcd for $C_{15}H_{19}N_4O_3$ [M + H]⁺ 303.1457, found 303.1457.

(S)-N-((1,4-Dioxooctahydropyridazino[1,2-a][1,2,4]-triazin-3-yl)methyl)-4-fluoro-benzamide (11b). Starting from 8 (20 mg, 0.089 mmol) and 4-fluorobenzoyl chloride (12 μL,

0,098 mmol), using general procedure B, **11b** was obtained as a white amorphous solid (22 mg, 0.069 mmol, 77%). R_f = 0.18 (DCM/MeOH, 9.6:0.4); $[\alpha]_D^{20}$ = -22.7 (c 0.18, MeOH); 1 H NMR (400 MHz, CDCl₃) δ 7.83 (dd, J = 8.1, 5.6 Hz, 2H), 7.38 (brt, J = 5.9 Hz, 1H), 7.07 (t, J = 8.4 Hz, 2H), 6.75 (s, 1H), 4.06 (brt, J = 5.0 Hz, 1H), 4.01-3.93 (m, 2H), 3.78-3.67 (m, 2H), 3.44-3.32 (m, 2H), 1.75-1.65 (m, 4H); 13 C NMR (100 MHz, CDCl₃) δ 167.4, 164.8 (d, J_{C-F} = 252 Hz), 163.6, 155.8, 130.0 (d, J_{C-F} = 3.0 Hz), 129.6 (d, J_{C-F} = 9.0 Hz, 2C), 115.5 (d, J_{C-F} = 21.9 Hz, 2C), 53.9, 44.7, 43.4, 40.3, 23.4, 22.9; 19 F NMR (376 MHz, CDCl₃) δ -107.7 (tt, J = 8.3, 5.6 Hz); HRMS (ESI-TOF): calcd for $C_{15}H_{18}N_4O_3F$ [M + H]⁺ 321.1363, found 321.1362.

(S)-N-((1,4-Dioxooctahydropyridazino[1,2-a][1,2,4]-triazin-3-yl)methyl)-3-methylbutan-amide (11c). Starting from 8 (30 mg, 0.134 mmol) and isovaleryl chloride (18 μL, 0,147 mmol), using general procedure B, 11c was obtained as a white amorphous solid (32 mg, 0.114 mmol, 85%). R_f = 0.10 (DCM/MeOH, 9.6:0.4); $[\alpha]_D^{20}$ = -28.2 (c 0.12, MeOH); 1 H NMR (400 MHz, CDCl₃) δ 6.30 (brt, J = 5.4 Hz, 1H), 5.96 (brs, 1H), 4.22-4.12 (m, 2H), 3.93 (t, J = 4.2 Hz, 1H), 3.91-3.84 (m, 1H), 3.52 (dt, J = 13.9, 4.7 Hz, 1H), 3.24-3.13 (m, 2H), 2.12-2.02 (m, 3H), 1.87-1.78 (m, 2H), 1.75-1.58 (m, 2H), 0.93 (d, J = 3.5 Hz, 6H); 13 C NMR (100 MHz, CDCl₃) δ 174.0, 163.9, 155.6, 53.9, 45.8, 44.5, 43.5, 38.6, 26.1, 23.4, 22.9, 22.4, 22.3; HRMS (ESI-TOF): calcd for $C_{13}H_{23}N_4O_3$ [M + H]+ 283.1770, found 283.1775.

(2S,5S)-tert-Butyl-(1-(((1,4-dioxooctahydropyridazino[1,2a][1,2,4]triazin-3-yl)methyl)-amino)-1-oxo-3-phenylpropan-2-yl)carbamate (11d). Starting from 8 (20 mg, 0.089 mmol) and Boc-L-phenylalanine (26 mg, 0.098 mmol), using general procedure C, 11d was obtained as a white amorphous solid (33 mg, 0.074 mmol, 84%). $R_f = 0.23$ (DCM/MeOH, 9.6:0.4); $[\alpha]_D^{20} = -16.6$ (c 0.13, MeOH); ¹H NMR (400 MHz, CDCl₃) δ 7.31–7.27 (m, 2H), 7.24–7.18 (m, 3H), 6.79 (brt, J = 5.7Hz, 1H), 5.88 (brs, 1H), 5.31 (brd, J = 8.2 Hz, 1H), 4.35–4.33 (m, 1H), 4.01–3.98 (m, 2H), 3.80–3.72 (m, 2H), 3.49–3.41 (m, 1H), 3.36–3.26 (m, 2H), 3.08–2.98 (m, 2H), 1.79–1.62 (m, 4H), 1.39 (s, 9H); 13 C NMR (100 MHz, CDCl₃) δ 172.7, 163.2, 155.4 (2C), 136.5, 129.3 (2C), 128.6 (2C), 126.9, 80.2, 56.0, 53.6, 44.7, 43.3, 39.3, 38.6, 28.3 (3C), 23.4, 22.9; HRMS (ESI-TOF): calcd for $C_{22}H_{32}N_5O_5[M + H]^+$ 446.2403, found 446.2396. A small quantity of the other diastereomer (2R,5S) was detected by HPLC at 220 nm, $t_R = 11.75 \text{ min}$ (C18 Waters Sunfire column (1 mL min⁻¹, 215 nm, linear gradient 5–100% acetonitrile in water in 20 min, 0.1% TFA)), diastereomeric ratio (dr) (2S,5S)/(2R,5S): 94:6.

(2S,5R)-tert-Butyl-(1-(((1,4-dioxooctahydropyridazino[1,2a][1,2,4]triazin-3-yl)methyl)-amino)-1-oxo-3-phenylpropan-2-yl)carbamate (11e). Starting from 8 (21 mg, 0.094 mmol) and Boc-D-phenylalanine (27 mg, 0.103 mmol), using general procedure C, 11e was obtained as a white amorphous solid (40 mg, 0.090 mmol, 96%). $R_f = 0.23$ (DCM/MeOH, 9.6:0.4); $[\alpha]_D^{20} = -29.8$ (c 0.10, MeOH); ¹H NMR (400 MHz, CDCl₃) δ 7.30–7.27 (m, 2H), 7.24–7.17 (m, 3H), 6.71 (brs, 1H), 5.98 (brs, 1H), 5.23 (brs, 1H), 4.35-4.34 (m, 1H), 4.06-4.02 (m, 2H), 3.80-3.72 (m, 2H), 3.42 (dt, J = 13.8, 5.8 Hz, 1H), 3.34-3.23 (m, 2H), 3.04 (brd, J = 4.9 Hz, 2H), 1.80-1.62 (m, 4H), 1.39 (s, 9H); 13 C NMR (100 MHz, CDCl₃) δ 172.7, 163.4, 155.5 (2C), 136.5, 129.3 (2C), 128.6 (2C), 126.9, 80.2, 56.0, 53.4, 44.6, 43.3, 39.3, 38.6, 28.2 (3C), 23.4, 22.9; HRMS (ESI-TOF): calcd for $C_{22}H_{32}N_5O_5[M + H]^+$ 446.2403, found 446.2392. A small quantity of the other diastereomer (2R,5R)

was detected by HPLC at 220 nm, $t_{\rm R}$ = 11.65 min (C18 Waters Sunfire column (1 mL min⁻¹, 215 nm, linear gradient 5–100% acetonitrile in water in 20 min, 0.1% TFA)), dr (2*S*,5*R*)/(2*R*,5*R*): 93:7.

tert-Butyl-(S)-((1,4-dioxooctahydropyridazino[1,2-a]-[1,2,4]triazin-3-yl)methyl)-carbamate (11f). To a solution of 8 (0.100 g, 0.446 mmol, 1 equiv) in dry MeOH (5 mL) was added 10 wt % Pd on carbon (10 mg) under argon. The resulting mixture was stirred at room temperature under a hydrogen atmosphere for 3 h. The reaction mixture was filtered over a pad of Celite, which was washed twiced with MeOH (2 × 10 mL). The filtrate was concentrated in vacuo and the resulting residue was straight away engaged in the next step. To a solution of the crude mixture in dry DCM (5 mL) was added Boc₂O (0.114 mL, 0.533 mmol, 1.2 equiv) at room temperature under argon. The reaction mixture was stirred at this temperature for 1 h under argon. The resulting mixture was diluted with DCM (20 mL) and H₂O (10 mL) and the aqueous layer was extracted three times with DCM (3 \times 15 mL). The combined organic layers were washed with brine (30 mL), dried over anhydrous Na₂SO₄, and concentrated in vacuo. The crude mixture was purified by silica gel chromatography (gradient 0-3% of MeOH in DCM) leading to 11f as a white amorphous solid (0.125 g, 0.419 mmol, 94%). $R_f = 0.16$ (DCM/MeOH, 9.6:0.4); $[\alpha]_D^{20} = -55.1$ (c 0.14, MeOH); ¹H NMR (400 MHz, DMSO- d_6) δ 7.18 (brs, 1H), 6.94 (brt, J = 6.0 Hz, 1H), 3.94–3.92 (m, 1H), 3.82–3.79 (m, 1H), 3.70 (td, J = 5.2, 2.1 Hz, 1H), 3.29 - 3.23 (m, 1H), 3.20 - $3.10 \text{ (m, 2H)}, 3.06 \text{ (dt, } J = 13.9, 5.3 \text{ Hz, 1H)}, 1.68-1.50 \text{ (m, } J = 13.9, 5.3 \text{ Hz, 1H)}, 1.68-1.50 \text{ (m, } J = 13.9, 5.3 \text{ Hz, } J = 1.50 \text{ (m, } J = 13.9, 5.3 \text{ Hz, } J = 1.50 \text{ (m, } J = 13.9, 5.3 \text{ Hz, } J = 1.50 \text{ (m, } J = 13.9, 5.3 \text{ Hz, } J = 1.50 \text{ (m, } J = 13.9, 5.3 \text{ Hz, } J = 1.50 \text{ (m, } J = 13.9, 5.3 \text{ Hz, } J = 1.50 \text{ (m, } J = 13.9, 5.3 \text{ Hz, } J = 1.50 \text{ (m, } J = 13.9, 5.3 \text{ Hz, } J = 1.50 \text{ (m, } J = 13.9, 5.3 \text{ Hz, } J = 1.50 \text{ (m, } J = 13.9, 5.3 \text{ Hz, } J = 1.50 \text{ (m, } J = 1.50 \text{ (m,$ 4H), 1.37 (s, 9H); 13 C NMR (100 MHz, DMSO- d_6) δ 162.3, 155.0, 153.9, 77.7, 53.4, 44.3, 42.0, 41.8, 27.7 (3C), 22.5, 21.9; HRMS (ESI-TOF): calcd for C₁₃H₂₃N₄O₄ [M + H]⁺ 299.1719, found 299.1713.

(S)-N-((1,4-Dioxo-2-(prop-2-yn-1-yl)octahydropyridazino-[1,2-a][1,2,4]triazin-3-yl)methyl)-benzamide (12a). Starting from 11a (14 mg, 0.046 mmol) and propargyl bromide (80 wt % in toluene, 26 μ L, 0.231 mmol), using general procedure D for 4 h, 12a was obtained as a white amorphous solid (12 mg, 0.035 mmol, 75%). $R_f = 0.23$ (*n*-pentane/EtOAc, 3:7); $[\alpha]_D^{20} =$ +8.1 (c 0.14, MeOH); ¹H NMR (400 MHz, CDCl₃) δ 7.77 (d, J = 7.5 Hz, 2H), 7.51 (t, J = 7.4 Hz, 1H), 7.44 (t, J = 7.4 Hz, 1Hz)2H), 6.70 (brs, 1H), 4.39 (d, I = 17.8 Hz, 1H), 4.30–4.27 (m, 2H), 4.17 (d, J = 17.8 Hz, 1H), 4.07 (brd, J = 12.7 Hz, 1H), 4.00 (brdt, J = 14.0, 5.1 Hz, 1H), 3.84 (dt, J = 13.6, 6.5 Hz, 1.00 (brdt, J = 14.0, 5.1 Hz, 1Hz)1H), 3.15-3.10 (m, 1H), 3.01 (ddd, I = 12.9, 10.2, 3.6 Hz, 1H), 2.37 (s, 1H), 1.80-1.64 (m, 4H); ¹³C NMR (100 MHz, CDCl₃) δ 167.6, 162.5, 153.8, 133.7, 131.8, 128.7 (2C), 127.0 (2C), 78.0, 73.8, 58.1, 46.2, 43.2, 39.0, 35.2, 23.5, 22.9; HRMS (ESI-TOF): calcd for $C_{18}H_{21}N_4O_3$ [M + H]⁺ 341.1614, found 341.1599.

(*S*)-*N*-((2-Butyl-1,4-dioxooctahydropyridazino[1,2-a]-[1,2,4]triazin-3-yl)methyl)-benzamide (12b). Starting from 11a (19 mg, 0.063 mmol) and 1-bromobutane (34 μ L, 0.314 mmol), using general procedure **D** for 24 h, 12b was obtained as a white amorphous solid (14 mg, 0.039 mmol, 62%). Three equivalents of BEMP and 1-bromobutane were added after 6 h. $R_f = 0.20$ (n-pentane/EtOAc, 1:4); $[\alpha]_D^{20} = +8.6$ (c 0.32, MeOH); ¹H NMR (400 MHz, CDCl₃) δ 7.78 (d, J = 7.5 Hz, 2H), 7.51 (t, J = 7.4 Hz, 1H), 7.43 (t, J = 7.5 Hz, 2H), 6.67 (brs, 1H), 4.43 (brd, J = 12.9 Hz, 1H), 4.28 (brd, J = 12.3 Hz, 1H), 4.00 (dd, J = 8.4, 4.7 Hz, 1H), 3.91–3.78 (m, 2H), 3.52 (ddd, J = 13.7, 8.3, 5.7, 1H), 3.01–2.89 (m, 3H), 1.84–1.67 (m, 3H), 1.62–1.56 (m, 3H), 1.32 (sxt, J = 7.1 Hz, 2H), 0.92

(t, J=7.2 Hz, 3H); 13 C NMR (100 MHz, CDCl₃) δ 167.6, 162.8, 154.1, 133.7, 131.8, 128.6 (2C), 127.0 (2C), 58.6, 46.0, 45.9, 43.1, 40.1, 30.1, 23.6, 23.0, 19.9, 13.7; HRMS (ESITOF): calcd for $C_{19}H_{27}N_4O_3$ [M + H]⁺ 359.2083, found 359.2064.

tert-Butyl-(S)-(3-(3-(benzamidomethyl)-1,4dioxohexahydropyridazino[1,2-a][1,2,4]-triazin-2(1H)-yl)propyl)carbamate (12c). Starting from 11a (20 mg, 0.066 mmol) and 3-(Boc-amino)propyl bromide (79 mg, 0.331 mmol), using general procedure D for 24 h, 12c was obtained as a white amorphous solid (19 mg, 0.041 mmol, 60%). Three equivalents of BEMP and 3-(Boc-amino) propyl bromide were added after 6 h. $R_f = 0.12$ (pentane/EtOAc, 1:4); $[\alpha]_D^{20} = +15.4$ (c 0.17, MeOH); ¹H NMR (400 MHz, CDCl₃) δ 7.79 (d, J =7.6 Hz, 2H), 7.52 (t, J = 7.5 Hz, 1H), 7.44 (t, J = 7.5 Hz, 2H), 6.77 (brs, 1H), 5.08 (brs, 1H), 4.41 (brd, J = 12.6 Hz, 1H), 4.22 (brd, J = 12.8 Hz, 1H), 4.00 (dd, J = 8.0, 4.4 Hz, 1H), 3.89 (dt, I = 13.4, 5.4 Hz, 1H), 3.79 - 3.72 (m, 1H), 3.60 - 3.53(m, 1H), 3.25-3.06 (m, 3H), 3.01-2.89 (m, 2H), 1.84-1.71 (m, 5H), 1.63-1.57 (m, 1H), 1.44 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 167.8, 162.6, 156.1, 154.4, 133.7, 131.8, 128.6 (2C), 127.0 (2C), 79.1, 59.3, 45.9, 43.8, 43.1, 40.3, 37.5, 28.7, 28.4 (3C), 23.5, 22.9; HRMS (ESI-TOF): calcd for $C_{23}H_{34}N_5O_5 [M + H]^+$ 460.2560, found 460.2543.

(S)-N-((2-Isobutyl-1,4-dioxooctahydropyridazino[1,2-a]-[1,2,4]triazin-3-yl)methyl)-benzamide (12d). Starting from 11a (30 mg, 0.099 mmol) and 1-bromo-2-methylpropane (54 μ L, 0.496 mmol), using general procedure D for 48 h, 12d was obtained as a white amorphous solid (4 mg, 0.039 mmol, 11%). Three equivalents of BEMP and 1-bromo-2-methylpropane were added after 6 h. $R_f = 0.23$ (EtOAc); $[\alpha]_D^{20} = +7.7$ (c 0.20, MeOH); ¹H NMR (400 MHz, CDCl₃) δ 7.78 (d, J =7.3 Hz, 2H), 7.52 (t, J = 7.3 Hz, 1H), 7.44 (t, J = 7.4 Hz, 2H), 6.58 (brs, 1H), 4.50 (brd, J = 12.7 Hz, 1H), 4.35 (br d, J = 12.6Hz, 1H), 3.99 (dd, J = 8.5, 4.9 Hz, 1H), 3.89-3.83 (m, 1H), 3.77 (dd, J = 13.8, 8.0 Hz, 1H), 3.52 (ddd, J = 13.6, 8.4, 5.6Hz, 1H), 2.95-2.85 (m, 2H), 2.69 (dd, J = 13.9, 7.0 Hz, 1H), 1.91 (sept, I = 6.8 Hz, 1H), 1.88–1.72 (m, 3H), 1.61–1.53 (m, 1H), 0.92 (d, J = 6.7 Hz, 3H), 0.88 (d, J = 6.7 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 167.7, 162.8, 154.3, 133.7, 131.9, 128.7 (2C), 127.0 (2C), 59.4, 53.3, 46.0, 43.1, 40.1, 27.3, 23.6, 23.0, 20.0, 19.7; HRMS (ESI-TOF): calcd for $C_{19}H_{27}N_4O_3$ [M + H]⁺ 359.2083, found 359.2080.

(S)-N-((2-(4-Fluorobenzyl)-1,4-dioxooctahydropyridazino-[1,2-a][1,2,4]triazin-3-yl)methyl)-benzamide (12e). Starting from 11a (30 mg, 0.099 mmol) and 4-fluorobenzyl bromide (62 μ L, 0.496 mmol), using general procedure D for 3 h, 12e was obtained as a white amorphous solid (32 mg, 0.078 mmol, 79%). $R_f = 0.24$ (n-pentane/EtOAc, 1:4); $[\alpha]_D^{20} = +39.9$ (c 0.27, MeOH); ¹H NMR (400 MHz, CDCl₃): δ 7.74 (d, J = 7.7 Hz, 2H), 7.50 (t, J = 7.5 Hz, 1H), 7.41 (t, J = 7.5 Hz, 2H), 7.30 (dd, J = 7.3, 5.8 Hz, 2H), 7.01 (t, J = 8.5 Hz, 2H), 6.65 (brt, J)= 4.8 Hz, 1H), 4.92 (d, J = 15.1 Hz, 1H), 4.41 (brd, J = 12.8)Hz, 1H), 4.28 (brd, J = 11.7 Hz, 1H), 4.24 (d, J = 15.1 Hz, 1H), 3.92 (dd, J = 7.7, 4.5 Hz, 1H), 3.70 (ddd, J = 13.8, 5.4, 4.9 Hz, 1H), 3.58 (ddd, J = 13.8, 7.4, 6.4 Hz, 1H), 2.98-2.91 (m, 2H), 1.83-1.66 (m, 3H), 1.61-1.51 (m, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 167.7, 162.5 (d, J_{C-F} = 247 Hz), 162.4, 154.2, 133.6, 132.0 (d, J_{C-F} = 3.2 Hz), 131.8, 130.1 (d, J_{C-F} = 8.1 Hz, 2C), 128.6 (2C), 127.0 (2C), 115.8 (d, J_{C-F} = 21.5 Hz, 2C), 58.0, 48.5, 46.3, 43.1, 39.5, 23.5, 22.9; ¹⁹F NMR (376 MHz, CDCl₃) δ -113.9 to -114.0 (m); HRMS (ESI-TOF): calcd for $C_{22}H_{24}N_4O_3F$ [M + H]⁺ 411.1832, found 411.1826.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acsomega.8b01752.

¹H NMR and ¹³C NMR spectra for all compounds (PDF)

AUTHOR INFORMATION

Corresponding Authors

*E-mail: nicolas.girard@unistra.fr. Phone: +33 (0)3 68 85 42 27 (N.G.).

*E-mail: dominique.bonnet@unistra.fr. Phone: + 33 (0)3 68 85 42 20 (D.B.).

ORCID ®

Marcel Hibert: 0000-0001-7786-7276 Nicolas Girard: 0000-0003-4610-9872 Dominique Bonnet: 0000-0002-8252-9199

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

This work was supported by the Centre National de la Recherche Scientifique (CNRS), the Université de Strasbourg (Unistra), and the Institut de Recherches Servier. We are grateful to Dr. Karolina Flidrova for NMR experiments and Sabine Lang for HRMS analysis (Service Commun d'Analyse, Unistra). The authors are also grateful to Prof. A. Ganesan (School of Pharmacy, University of East Anglia, U.K.) for fruitful discussions.

REFERENCES

- (1) Stockwell, B. R. Exploring biology with small organic molecules. *Nature* **2004**, 432, 846–854.
- (2) Lovering, F.; Bikker, J.; Humblet, C. Escape from flatland: increasing saturation as an approach to improving clinical success. *J. Med. Chem.* **2009**, *52*, *6752*–*6756*.
- (3) Borthwick, A. D. 2,5-Diketopiperazines: synthesis, reactions, medicinal chemistry, and bioactive natural products. *Chem. Rev.* **2012**, 112, 3641–3716.
- (4) (a) Hoffman, R. V.; Nayyar, N. Reaction of hydrazines with.alpha.-lactams for the preparation of 1,2,4-triazine-3,6-diones and aza-urea peptide mimetics. *J. Org. Chem.* **1995**, *60*, 5992–5994. (b) Hoffman, R. V.; Reddy, M. M.; Klumas, C. M.; Cervantes-Lee, F. The reactions of hydrazines with α -lactams. Regiochemistry of hydrazine addition and subsequent ring closure to *N*-aminohydantoins or 1,2,4-triazine-3,6-diones. *J. Org. Chem.* **1998**, *63*, 9128–9130.
- (5) Obreza, A.; Urleb, U. A two-step synthesis of hexahydropyrrolo-[1,2-d][1,2,4]triazine-1,4-dione and related compounds. *Synth. Commun.* **2003**, 33, 1011–1018.
- (6) Bourguet, C. B.; Sabatino, D.; Proulx, C.; Klocek, S.; Lubell, W. D. Solution-phase submonomer diversification of aza-dipeptide building blocks and their Application in aza-peptide and aza-DKP synthesis. *J. Pept. Sci.* **2010**, *16*, 284–296.
- (7) Ivanovich, R. A.; Vincent-Rocan, J.-F.; Elkaeed, E. B.; Beauchemin, A. M. One-pot synthesis of aza-diketopiperazines enabled by controlled reactivity of *N*-isocyanate precursors. *Org. Lett.* **2015**, *17*, 4898–4901.
- (8) Bonnet, D.; Margathe, J.-F.; Radford, S.; Pflimlin, E.; Riche, S.; Doman, P.; Hibert, M.; Ganesan, A. Combinatorial aid for underprivileged scaffolds: solution and solid-phase strategies for a rapid and efficient access to novel aza-diketopiperazines (aza-DKP). ACS Comb. Sci. 2012, 14, 323–334.

- (9) (a) Regenass, P.; Margathe, J.-F.; Mann, A.; Suffert, J.; Hibert, M.; Girard, N.; Bonnet, D. Diastereoselective synthesis of novel azadiketopiperazines via a domino cyclohydro-carbonylation/addition process. *Chem. Commun.* **2014**, *50*, 9657–9660. (b) Regenass, P.; Riche, S.; Peron, F.; Rognan, D.; Hibert, M.; Girard, N.; Bonnet, D. A step-economical multicomponent synthesis of 3D-shaped azadiketopiperazines and their drug-like chemical space analysis. *Org. Biomol. Chem.* **2016**, *14*, 8859–8863.
- (10) Regenass, P.; Bosc, D.; Riche, S.; Gizzi, P.; Hibert, M.; Karmazin, L.; Ganesan, A.; Bonnet, D. Comparative study of the synthesis and structural and physicochemical properties of diketopiperazines vs aza-diketopiperazines. *J. Org. Chem.* **2017**, *82*, 3239–3244.
- (11) Wauters, I.; Goosens, H.; Delbeke, E.; Muylaert, K.; Roman, B. I.; Van Hecke, K.; Van Speybroeck, V.; Stevens, C. V. Beyond the diketopiperazine family with alternatively bridged brevianamide F analogues. *J. Org. Chem.* **2015**, *80*, 8046–8054.
- (12) Sakamoto, H.; Egashira, S.; Saito, N.; Kirisako, T.; Miller, S.; Sasaki, Y.; Matsumoto, T.; Shimonishi, M.; Komatsu, T.; Terai, T.; Ueno, T.; Hanaoka, K.; Kojima, H.; Okabe, T.; Wakatsuki, S.; Iwai, K.; Nagano, T. Gliotoxin suppresses NF-κB activation by selectively inhibiting linear ubiquitin chain assembly complex (LUBAC). ACS Chem. Biol. 2015, 10, 675–681.
- (13) Jida, M.; Tourwé, D.; Ballet, S. Highly stereoselective one-pot construction of trisubstituted tetrahydro-β-carboline-fused diketopiperazines: a synthetic route towards cialis analogues. RSC Adv. 2014, 4, 38159–38163.
- (14) Colombo, R.; Mingozzi, M.; Belvisi, L.; Arosio, D.; Piarulli, U.; Carenini, N.; Perego, P.; Zaffaroni, N.; De Cesare, M.; Castiglioni, V.; Scanziani, E.; Gennari, C. Synthesis and biological evaluation (in vitro and in vivo) of cyclic arginine—glycine—aspartate (RGD) peptidomimetic—paclitaxel conjugates targeting integrin $\alpha_V \beta_3$. *J. Med. Chem.* **2012**, *55*, 10460–10474.
- (15) Kim, J.; Movassaghi, M. General approach to epipolythiodiketopiperazine alkaloids: total synthesis of (+)-chaetocins A and C and (+)-12,12'-dideoxychetracin A. J. Am. Chem. Soc. 2010, 132, 14376–14378.
- (16) Doro, F.; Colombo, C.; Alberti, C.; Arosio, D.; Belvisi, L.; Casagrande, C.; Fanelli, R.; Manzoni, L.; Parisini, E.; Piarulli, U.; Luison, E.; Figini, M.; Tomassetti, A.; Civera, M. Computational design of novel peptidomimetic inhibitors of cadherin homophilic interactions. *Org. Biomol. Chem.* **2015**, *13*, 2570–2573.
- (17) (a) Kolb, H. C.; Sharpless, K. B. The growing impact of click chemistry on drug discovery. *Drug Discovery Today* **2003**, *8*, 1128–1137. (b) Hou, J.; Liu, X.; Shen, J.; Zhao, G.; Wang, P. G. The impact of click chemistry in medicinal chemistry. *Expert Opin. Drug Discovery* **2012**, *7*, 489–501. (c) Thirumurugan, P.; Matosiuk, D.; Jozwiak, K. Click chemistry for drug development and diverse chemical—biology applications. *Chem. Rev.* **2013**, *113*, 4905–4979.
- (18) Wang, J.; Liang, Y.-L.; Qu, J. Boiling water-catalyzed neutral and selective N-Boc deprotection. *Chem. Commun.* **2009**, 5144–5146.
- (19) Cohen, S. B.; Halcomb, R. L. Application of serine- and threonine-derived cyclic sulfamidates for the preparation of S-linked glycosyl amino acids in solution- and solid-phase peptide synthesis. J. Am. Chem. Soc. 2002, 124, 2534–2543.
- (20) Bouzide, A.; Sauvé, G. Highly selective silver(I) oxide mediated monoprotection of symmetrical diols. *Tetrahedron Lett.* **1997**, 38, 5945–5948.
- (21) Zhu, Y.; van der Donk, W. A. Convergent synthesis of peptide conjugates using dehydroalanines for chemoselective ligations. *Org. Lett.* **2001**, *3*, 1189–1192.
- (22) Tullberg, M.; Grøtli, M.; Luthman, K. Synthesis of functionalized, unsymmetrical 1,3,4,6-tetrasubstituted 2,5-diketopiperazines. *J. Org. Chem.* **2007**, 72, 195–199.
- (23) Zhao, C.; Sham, H. L.; Sun, M.; Stoll, V. S.; Stewart, K. D.; Lin, S.; Mo, H.; Vasavanonda, S.; Saldivar, A.; Park, C.; McDonald, E. J.; Marsh, K. C.; Klein, L. L.; Kempf, D. J.; Norbeck, D. W. Synthesis and activity of *N*-acyl azacyclic urea HIV-1 protease inhibitors with high

potency against multiple drug resistant viral strains. *Bioorg. Med. Chem. Lett.* **2005**, *15*, 5499–5503. (24) Cuny, G. D.; Buchwald, S. L. Practical, high-yield,

- (24) Cuny, G. D.; Buchwald, S. L. Practical, high-yield, regioselective, rhodium-catalyzed hydroformylation of functionalized.alpha.-olefins. *J. Am. Chem. Soc.* **1993**, *115*, 2066.
- (25) Ma, G.; Nguyen, H.; Romo, D. Concise total synthesis of (\pm) -salinosporamide A, (\pm) -cinnabaramide A, and derivatives via a bis-cyclization process: implications for a biosynthetic pathway? *Org. Lett.* **2007**, *9*, 2143–2146.